Melanocortin receptor-mediated mobilization of intracellular free calcium in HEK293 cells

KATHLEEN G. MOUNTJOY,1 PHILIP L. KONG,1 JOHN A. TAYLOR,2 DERRIL H. WILLARD,3 AND WILLIAM O. WILKISON3

1Research Centre for Developmental Medicine and Biology, Department of Paediatrics, and 2School of Biological Sciences, University of Auckland, Auckland 1, New Zealand; and 3Glaxo Wellcome Research Institute, Research Triangle Park, North Carolina 27709

Received 7 October 1999; accepted in final form 21 November 2000

Mountjoy, Kathleen G., Philip L. Kong, John A. Taylor, Derril H. Willard, and William O. Wilkinson. Melanocortin receptor-mediated mobilization of intracellular free calcium in HEK293 cells. Physiol Genomics 5: 11–19, 2001.—Mouse melanocortin receptors, MC1-R, MC3-R, MC4-R, and MC5-R, when expressed in HEK293 cells and stimulated with either α-melanocyte-stimulating hormone (α-MSH) or desacetyl-α-MSH, mediate increases in intracellular free calcium concentration ([Ca2+]i) with EC50 values between 0.3 and 4.3 nM. The increase in [Ca2+]i is cholera toxin sensitive and pertussis toxin insensitive. The mechanism involves calcium mobilization from intracellular stores without a transient rise in inositol trisphosphate. Mouse agouti protein (55 nM) is a competitive antagonist of the mouse MC1-R (mMC1-R) to melanocortin peptides (8-fold), coupling the mMC1-R to increased [Ca2+]i. Agouti protein (55 nM) significantly increased the EC50 for α-MSH (3-fold), and 550 nM agouti protein significantly increased the EC50 for desacetyl-α-MSH (4-fold), coupling the mMC4-R to a rise in [Ca2+]i. However, agouti protein antagonism of the MC4-R may not be competitive since there was a trend for the maximum response to also increase. There was no significant antagonism of the MC3-R and MC5-R by agouti protein (55 nM). Understanding the physiological relevance of the transduction of a calcium signal by melanocortin peptides may be important for future development of therapeutic targeting of the melanocortin receptors.

A FAMILY OF FIVE MELANOCORTIN receptors (MC1-R to MC5-R), through which peptides derived from proopiomelanocortin function, have been identified. The MC1-R is expressed in melanocytes and plays a role in the pigmentation response (26). The MC2-R is expressed in adrenal cortex and is involved in the stress response (26). Localization of MC3-R (33) and MC4-R (25) mRNA to regions in adult rat brain indicated both receptors may function in neuroendocrine, cardiovascular, and food and water intake regulation. Targeted disruption of the gene encoding the MC4-R identified a role for this receptor in appetite control, weight regulation, and regulation of linear growth (15). The MC4-R is probably also involved in autonomic control and higher learning processes. The MC5-R regulates exocrine gland function (4).

Overexpression of G protein-coupled receptors in heterologous cell lines is widely used to begin to understand ligand-induced intracellular signaling pathways. Numerous studies have confirmed that all members of the melanocortin receptor family couple to the adenylyl cyclase/protein kinase A (PKA) intracellular signaling pathway when expressed in heterologous cells (10–12, 19, 25, 26, 33). Mouse agouti protein is a competitive antagonist of the mouse MC1-R (mMC1-R) coupling to the PKA signaling pathway and thereby regulates pigmentation (21). Agouti protein also antagonizes the MC4-R coupling to the PKA signaling pathway (21), but in this case agouti protein decreases both the sensitivity and maximum responsiveness of the mouse MC4-R (mMC4-R) to melanocortin peptides (27). Agouti protein antagonism of MC4-R underlies the obesity and non-insulin-dependent diabetes mellitus (NIDDM) phenotype in mice ectopically expressing agouti protein.

A second melanocortin receptor-mediated intracellular signaling pathway, at least for the MC3-R, has also been described (18). Konda et al. (18) showed that low concentrations of melanocortin peptides stimulate a transient rise in inositol trisphosphate (IP3), and when the PKA pathway is inhibited, high concentrations of melanocortin peptides increase intracellular calcium concentration ([Ca2+]i) in heterologous cells. Very little else is known about melanocortin peptides coupling their receptors to signaling pathways resulting in changes in [Ca2+]i. Agouti protein, however, is associated with changes in intracellular calcium. Agouti protein induces a slow rise in [Ca2+]i in heterologous cells overexpressing melanocortin receptors (17) and also in isolated skeletal muscle myocytes from wild-type mice (38), differentiated 3T3-L1 adipocytes (16), and the pancreatic β-cell (37). A link between agouti protein, melanocortin receptors, and calcium signaling was strengthened by the finding that [Ca2+]i is elevated in
skeletal muscle in A^v yellow mice compared with non-agouti (a/ata) mice (38).

To further understand the functions of melanocortin peptides and their receptors, in particular the mechanism by which the MC4-R influences weight regulation, as well as the mechanism by which ectopic agouti protein drives obesity, it is necessary to know whether the melanocortin receptors couple to a signaling pathway that results in alterations of [Ca^{2+}], and whether agouti protein can influence this pathway. We have therefore investigated two naturally occurring melanocortin peptides coupling four different melanocortin receptors to a signaling pathway in HEK293 cells that results in elevation of [Ca^{2+}], and we tested for antagonism of this pathway by agouti protein.

METHODS

The melanocortin peptides, desacetyl-a-melanocyte-stimulating hormone (desacetyl-o-MSH) and a-MSH, were purchased from Bachem California. The acetoxymethyl ester of fura-2 (fura-2/AM) and the ionophore, A23187, were purchased from Molecular Probes (Eugene, OR). The production of recombinant mouse agouti protein has previously been described (36). Nifedipine and diltiazem were Sigma products and a gift from Dr. D. R. Mason. [3H]inositol and [5,6,8,9,11,12,14,15-3H(N)]arachidonic acid were purchased from Amersham Life Science (Buckinghamshire, UK). Pertussis toxin and forskolin were purchased from Sigma (St. Louis, MO), and cholera toxin and the Rp isomer of adenosine 3',5'-cyclic monophosphorothioate (Rp-cAMPS) were purchased from Calbiochem.

Cell lines. Stably transfected human embryonic kidney HEK293 cells expressing the mMC1-R have previously been described (26). The coding sequences of mMC3-R and mMC4-R were obtained by probing a mouse genomic library, 129 strain (Stratagene), with the corresponding rat receptor DNA sequences. The mMC5-R cDNA was obtained from brown fat cDNA using RT-PCR with primers specific for this gene (19). DNA coding sequences of the mouse melanocortin receptors (mMC3, mMC4, and mMC5) were subcloned into pcDNAIneo vector (Invitrogen), and these were stably transfected into HEK293 cells using a modified calcium phosphate procedure (3). Stably transfected cells were selected in DMEM containing 10% newborn calf serum (NCS) and 1 mg/ml G418, then grown with 500 μg/ml G418.

Measurement of inositol phosphates. Inositol phosphates were measured according to a previously published method (29). Cells were plated into 24-well plates at a density of 10^5 cells per well, and on day 3 the media was aspirated and the cells were prelabeled with 0.7 μCi myo-[2-3H]inositol per well in 250 μl DMEM, 10% NCS for 48 h at 37°C. Cells were then washed with 1 ml PBS, warmed to 37°C, and incubated in 250 μl of 0.1% BSA-DMEM containing 10 mM LiCl for 30 min at 37°C. This media was then aspirated, and the cells were incubated with or without agonists in the same medium for 20 min at 37°C. The reaction was stopped by the addition of 250 μl ice-cold 10% trichloroacetic acid per well, and the cells were scraped and transferred to Eppendorf tubes on ice. Following vortexing and centrifugation, the cell extracts were transferred to clean Eppendorf tubes and desalted by the addition of 600 μl of a 1:1 mix of 1,1,2-trichlorotrifluoroethane/tri-n-octylamine mixture. The upper phase was removed, and inositol phosphates were separated by ion exchange using 0.5-ml columns of Dowex AG 1-X8 formate form (Bio-Rad) ion-exchange resin. Prior to adding the samples, the columns were washed with 5 ml water. Following the addition of the samples, the columns were washed with 7.5 ml of 5 mM myo-inositol in water. Total inositol phosphates were eluted by adding 1.5 ml of 1 M ammonium formate/0.1 M formic acid and then counted using liquid scintillation spectrometry in a beta scintillation counter (LKB 1219 Rack-Beta).

Measurement of [Ca^{2+}]. Confluent cells from a T75 culture flask were washed briefly with versene and then detached from the plates by incubation with 2 ml versene at 37°C for 3 min. The cells were diluted in 5 ml DMEM containing 10% NCS and pelleted by centrifugation. The cells were resuspended in DMEM containing 0.1% BSA and 25 mM HEPES, then loaded with fura-2/AM (2 μM) using gentle rocking for 20 min at 37°C. After loading, the cells were pelleted by centrifugation and washed two times with calcium loading buffer (CLB) (composition, in mM: 132 NaCl, 5 KCl, 5 Na2HPO4, 1.2 NaH2PO4, 2H2O, 1 CaCl2, 0.8 MgCl2, and 1 glucose). Cells from each T75 flask were finally resuspended in 12 ml CLB and maintained at room temperature while experiments were carried out on 1.8-ml aliquots at 37°C in a Hitachi model F-4500 dual-wavelength spectrophotometer equipped with a thermostated cuvette holder, using previously described methods (32). The excitation wavelength was changed once per second between 340 and 380 nm, and emission was recorded at 510 nm. Fluorescence ratios R = F530/F500 were recorded, and [Ca^{2+}], was calculated using the equation of Grynkiewicz et al. (14), with K_d = 224. Values for R_min and R_max were determined for each experiment by successive addition of ionomycin (20 μl of 1 mM stock in methanol) and EGTA (20 μl of 0.5 M stock in water).

Quenching of fura-2 fluorescence by Mn^{2+} was measured by exciting the dye at 360 nm, where emission measured at 510 nm is independent of ambient calcium concentration. This technique detects agonist-stimulated opening of cation channels in different cell types (9). Thapsigargin was used to test for the release of intracellular calcium. A stock solution of thapsigargin (1 μM) dissolved in dimethyl sulfoxide (DMSO) was prepared. The rapidly exchangeable calcium pool of the endoplasmic reticulum was empty ~150 s following the addition of thapsigargin to the cuvette (final concentration of 10^{-6} M). The cells were then tested for responses to 10^{-6} M α-MSH or 10^{-3} M carbachol.

Representatives of two chemically distinct classes of organic antagonists of L-type voltage-sensitive Ca^{2+} channels (nifedipine and diltiazem) and two inorganic blocking ions (Cd^{2+} and La^{3+}) were used to test whether melanocortin peptides stimulate the entry into cells of extracellular calcium through calcium channels. Stock solutions containing the channel blockers were prepared on the day of each experiment. Nifedipine (2 μM) was dissolved in DMSO, whereas diltiazem (5 × 10^{-3} M), Cd(NO3)2 (0.5 M), and La(NO3)3 (0.5 M) were dissolved in water. Calcium channel blockers were added to the cuvette ~50 s before the addition of 10^{-6} M α-MSH.

Sensitivity to cholera and pertussis toxin were used to test for G protein-mediated mobilization of [Ca^{2+}], by α-MSH, whereas forskolin and Rp-cAMPS, a competitive antagonist of PKA, were used to test for PKA-mediated mobilization of [Ca^{2+}]. Cholera and pertussis toxin were dissolved in DMEM and added to cells for 16 h at final concentrations of 2 μg/ml and 100 ng/ml, respectively. Forskolin (10 mM) was dissolved in DMSO. Rp-cAMPS (5 mM) was dissolved in DMEM containing 0.1% BSA and added to the fura-2-preloaded cells 15–45 min prior to testing the cells response for mobilization of [Ca^{2+}].

http://physiolgenomics.physiology.org
Measurement of agouti antagonism of melanocortin peptide-stimulated increases in $[\text{Ca}^{2+}]_i$. Dose response curves were performed pairwise: one without agouti protein and one in the presence of agouti protein. Melanocortin peptides ($10^{-10}$, $10^{-9}$, $10^{-8}$, $10^{-7}$, and $10^{-6}$ M) and agouti protein (10, 55, and 550 nM) were diluted DMEM containing 0.1% BSA and 25 mM HEPES. Agouti protein was added 150 s prior to the addition of either $\alpha$-MSH or desacetyl-$\alpha$-MSH. The change in $[\text{Ca}^{2+}]_i$ for each dose was determined after taking the means of four to seven points at both the baseline and peak responses. The KALEIDAGRAPH software package (Synergy Software, Reading, PA) was used for fitting curves to the data and calculating EC$_{50}$ and maximum response values. The R values for all curves were $>0.8$.

Statistics. Comparisons between groups were made using Sigmastat (Jandel Scientific) and either one-way ANOVA or the Student’s $t$-test.

RESULTS

$\alpha$-MSH and desacetyl-$\alpha$-MSH stimulate increases in $[\text{Ca}^{2+}]_i$. $\alpha$-MSH induced a transient rise in $[\text{Ca}^{2+}]_i$ in HEK293 cells expressing mMC1, mMC3, mMC4, and mMC5 receptors but not in untransfected HEK293. This increase was dose dependent, with $10^{-10}$ M $\alpha$-MSH having a small effect and a maximum response observed with by $10^{-7}$–$10^{-6}$ M $\alpha$-MSH (Fig. 1). Desacetyl-$\alpha$-MSH also coupled each of these receptors to rises in $[\text{Ca}^{2+}]_i$. EC$_{50}$ values for the increase in $[\text{Ca}^{2+}]_i$ induced by both $\alpha$-MSH and desacetyl-$\alpha$-MSH are given in Table 1. Carbachol, through stimulation of endogenous muscarinic receptors in HEK293 cells, also induced a dose-dependent increase in $[\text{Ca}^{2+}]_i$ at con-

![Image of Figure 1](http://physiolgenomics.physiology.org)

This increase was dose dependent, with $10^{-10}$ M $\alpha$-MSH having a small effect and a maximum response observed with by $10^{-7}$–$10^{-6}$ M $\alpha$-MSH (Fig. 1). Desacetyl-$\alpha$-MSH also coupled each of these receptors to rises in $[\text{Ca}^{2+}]_i$. EC$_{50}$ values for the increase in $[\text{Ca}^{2+}]_i$ induced by both $\alpha$-MSH and desacetyl-$\alpha$-MSH are given in Table 1. Carbachol, through stimulation of endogenous muscarinic receptors in HEK293 cells, also induced a dose-dependent increase in $[\text{Ca}^{2+}]_i$ at con-

Table 1. EC$_{50}$ values for melanocortin peptides inducing an increase in mobilization of intracellular calcium in HEK293 cells stably expressing the mouse melanocortin receptors

<table>
<thead>
<tr>
<th>Receptor</th>
<th>EC$_{50}$, nM</th>
</tr>
</thead>
<tbody>
<tr>
<td>MC1-R</td>
<td></td>
</tr>
<tr>
<td>MC3-R</td>
<td></td>
</tr>
<tr>
<td>MC4-R</td>
<td></td>
</tr>
<tr>
<td>MC5-R</td>
<td></td>
</tr>
</tbody>
</table>

$\alpha$-MSH, 4.3 ± 1.6 (5) 1.3 ± 0.6 (4) 0.5 ± 0.3 (15) 1.3 ± 0.7 (4)
desacetyl-$\alpha$-MSH, 2.7 ± 1.9 (6) 1.3 ± 0.3 (4) 0.3 ± 0.1 (9) 1.6 ± 0.8 (4)

Values are means ± SD; numbers in parentheses are numbers of dose-response curves. $\alpha$-MSH, $\alpha$-melanocyte stimulating hormone.
Concentrations as low as $10^{-6}$ M with a peak response observed with $10^{-3}$ M (Fig. 1). Carbachol induced a classic IP$_3$-mediated rise in [Ca$^{2+}$]$_i$ characterized by a rapid increase, reflecting release from intracellular stores, followed by a subsequent decline. In contrast, the melanocortin peptide stimulation of [Ca$^{2+}$]$_i$ exhibited a slower increase that was sustained following the maximum response. Examination of inositol phosphate levels in cells following exposure to melanocortin peptides revealed the absence of a detectable increase (Fig. 2).

α-MSH stimulates release of calcium from intracellular stores. The observed rise in [Ca$^{2+}$]$_i$ (Fig. 1) could be due to release of Ca$^{2+}$ from intracellular stores or reflect an increase in Ca$^{2+}$ influx via plasma membrane channels. Three approaches were taken to distinguish the source of increased cytoplasmic Ca$^{2+}$ in response to melanocortin peptides. First, the Ca$^{2+}$-ATPase inhibitor, thapsigargin, was used to empty the rapidly exchangeable calcium pool of the endoplasmic reticulum (22). Pretreatment of HEK293 cells with 1 μM thapsigargin abolished a rise in [Ca$^{2+}$]$_i$ in response to either α-MSH ($10^{-6}$ M) or carbachol ($10^{-3}$ M) (Fig. 3). Second, we tested the possibility that α-MSH could open Ca$^{2+}$ permeable cation channels on the plasma membrane through which Mn$^{2+}$ (a calcium surrogate) can also enter (22). In the presence of external Mn$^{2+}$, neither α-MSH nor carbachol induced a decrease in fura-2 fluorescence at its isobestic wavelength (360 nm), whereas each one induced a transient increase in fura-2 fluorescence at 340 nm (Fig. 4). In contrast, ionomycin caused a decrease in fura-2 fluorescence at 360 nm, consistent with quenching of the dye due to Mn$^{2+}$ influx. Third, we tested whether α-MSH caused Ca$^{2+}$ influx through L-type voltage-gated calcium channels in the plasma membrane using antagonist blocking ions, Cd$^{2+}$ and La$^{3+}$ (1 mM), as well as the organic antagonists, nifedipine (10 μM) and diltiazem (100 μM). It is unclear why the basal calcium fluctuated for two of these blockers. Importantly, however, all four calcium channel blockers failed to reduce the α-MSH-induced rise of [Ca$^{2+}$]$_i$, nor did they have any effect on the sustained elevation of calcium levels (Fig. 5).

α-MSH-induced mobilization of [Ca$^{2+}$]$_i$ is cholera toxin sensitive. Cholera toxin (2 μg/ml) significantly attenuated $10^{-6}$ M α-MSH-induced mobilization of [Ca$^{2+}$]$_i$ in mMC4-R-expressing cells from 105 ± 10 nM (mean ± SD, n = 3) to 20 ± 7 nM (n = 3, P < 0.0003).
Pertussis toxin (100 ng/ml) had no effect on α-MSH-induced mobilization of \([Ca^{2+}]_i\) in these cells (no pretreatment, 77 ± 32 nM; and 109 ± 25 nM for pertussis toxin treatment; means ± SD, \(n = 3\)).

Rp-cAMPS (5 × 10⁻⁵ M) appeared to slightly attenuate 10⁻⁸ M α-MSH-induced mobilization of \([Ca^{2+}]_i\) in mMC4-R-expressing cells (no treatment, 75 ± 5 nM; and Rp-cAMPS treatment, 66 ± 26 nM; means ± SD, \(n = 3\)), but this was not significant. This dose of Rp-cAMPS, however, significantly attenuated 10⁻⁸ M forskolin-induced mobilization of \([Ca^{2+}]_i\) (no treatment, 36 ± 9 nM; and Rp-cAMPS treatment, 18 ± 2 nM; means ± SD, \(n = 3\), \(P < 0.02\)).

Mouse agouti protein influences the α-MSH and desacetyl-α-MSH-induced increases in \([Ca^{2+}]_i\) through the mMC1 and mMC4 receptors but not through the mMC3 or mMC5 receptors. Mouse agouti protein (55–550 nM) significantly decreased the sensitivity of α-MSH-mediated increased \([Ca^{2+}]_i\) via mMC4-R (Fig. 6). A relatively high concentration of agouti protein (550 nM) also significantly increased the EC₅₀ for desacetyl-α-MSH coupling the mMC4-R to mobilization of \([Ca^{2+}]_i\). The \(R_{max}\) values achieved for both α-MSH and desacetyl-α-MSH in the presence of agouti protein, however, appeared to increase but were not significantly different from that achieved in the absence of agouti protein (Fig. 6). The sensitivity of α-MSH and desacetyl-α-MSH coupling of the mMC4-R to the \([Ca^{2+}]_i\) pathway was decreased up to 12- and 4-fold, respectively, by 550 nM mouse agouti protein (Fig. 6). The trend for agouti protein to increase the \(R_{max}\) would contribute to the agouti-induced decrease in sensitivity and suggests that agouti protein may not be a competitive antagonist of the mMC4-R, but it influences, and possibly enhances, the ability of melanocortin peptides to couple the MC4-R to the calcium signaling pathway.

Mouse agouti protein (55 nM) antagonized α-MSH-mediated and desacetyl-α-MSH-mediated increases in \([Ca^{2+}]_i\) via mMC1-R (Fig. 7). This antagonism, however, exhibited competitive kinetics, since the \(R_{max}\) achieved in the presence of agouti protein was similar to that observed in the absence of agouti protein.
Fig. 6. Agouti protein influences α-MSH and desacetyl-α-MSH coupling the mMC4-R to mobilization of [Ca^{2+}]_{i}. Dose response curves for measuring [Ca^{2+}]_{i}, using α-MSH or desacetyl-α-MSH were performed in the absence or presence of 10, 55, and 550 nM mouse agouti protein, and the resulting curves were analyzed using KALEIDAGRAPH software. EC_{50} (A) and maximum response (B) data points are means ± SD of multiple separate experiments. The number of experiments is shown in parentheses for each point. Significant differences between the EC_{50} values generated in the absence or presence of agouti protein were determined using one-way ANOVA. *P < 0.05.

![Diagram A](image1)

![Diagram B](image2)

to that achieved in the absence of agouti. The sensitivity of α-MSH and desacetyl-α-MSH coupling the mMC1-R to the [Ca^{2+}]_{i} pathway was significantly decreased by six- and eightfold, respectively, by 55 nM mouse agouti protein (Fig. 7). Mouse agouti protein did not significantly antagonize α-MSH or desacetyl-α-MSH coupling the mMC3 and mMC5 receptors to the [Ca^{2+}]_{i} pathway (Fig. 7).

Mouse agouti protein alone, or DMEM containing 0.1% BSA and 25 mM HEPES alone, induced a small increase in [Ca^{2+}]_{i} (<30 nM) that was independent of the agouti dose.

**DISCUSSION**

We show that mouse MC1, MC3, MC4, and MC5 receptors when stably expressed in HEK293 cells and stimulated with either α-MSH or desacetyl-α-MSH, mediate increases in [Ca^{2+}]_{i} in addition to activating adenyl cyclase. Desacetyl-α-MSH is more abundant than α-MSH in the brain (5, 28), the fetus (8, 30), human blood (7), and amniotic fluid (23), and the functional responses to desacetyl-α-MSH and α-MSH differ in vivo. For example, subcutaneous injections of desacetyl-α-MSH do not affect pigmentation, and desacetyl-α-MSH is more potent at stimulating growth than is α-MSH (24, 35). There is, however, little information on desacetyl-α-MSH interactions with melanocortin receptors in vitro. In this study, we have compared and contrasted the ability of desacetyl-α-MSH and α-MSH to couple melanocortin receptors to a calcium mobilization signaling pathway. In contrast to the different in vivo effects of these peptides, desacetyl-α-MSH and α-MSH similarly activate MC1, MC3, MC4, and MC5 receptors coupling to the calcium signaling pathway with EC_{50} values between 0.3 and 4.3 nM.

The increase in [Ca^{2+}]_{i} is cholera toxin sensitive and pertussis toxin insensitive and is due to mobilization from intracellular stores by a mechanism that does not involve a transient increase in IP_{3}. The absence of increased [IP_{3}], despite a clear increase in [Ca^{2+}]_{i} in response to α-MSH, suggests the involvement of an alternative second messenger. The parathyroid hormone (34) and dopamine D_{1} (20) receptors expressed in HEK293 cells also couple to an intracellular signaling pathway that does not involve IP_{3}, but results in Ca^{2+} release from intracellular stores (34). Interestingly, forskolin increases [Ca^{2+}]_{i} in HEK293 cells, implicating cAMP as the second messenger that mobilizes calcium in these cells (21), and since the forskolin response is attenuated by prestimulation with carbachol, the cAMP-dependent store is considered part of the IP_{3}-sensitive calcium pool (20). In hepatocytes, cAMP has also been shown to sensitize IP_{3}-mediated release of calcium stores (2). Our studies, however, do not provide strong evidence for the involvement of cAMP as a second messenger for α-MSH-induced mobilization of calcium. First, prestimulation with carbachol (10^{-4} M) did not affect the calcium response to α-MSH (10^{-8} M) (data not shown), and second, 5 \times 10^{-5} M Rp-cAMPS did not significantly attenuate α-MSH-mediated release (10^{-8} M) calcium mobilization in HEK293 cells.

In addition to the intracellular Ca^{2+} release channels (IP_{3} and ryanodine receptors), Ca^{2+} can be mobilized by arachidonic acid, a product of phospholipase A_{2} (PLA_{2}) activity (1), and also by a novel Ca^{2+} release mechanism activated by nicotinic acid adenine dinucleotide phosphate (NAADP) (6, 13). In this study, we were unable to show any melanocortin peptide-stimulated increase of [^{3}H]arachidonic acid (data not shown), which suggests that melanocortin receptors do not couple to PLA_{2} in HEK293 cells. Furthermore, it is unlikely that NAADP is the signaling pathway that melanocortin receptors use to mobilize Ca^{2+}, since NAADP mobilizes this ion from a pool distinct from the endoplasmic reticulum.

A similar response to that reported in our study, namely, a rapid transient elevation of [Ca^{2+}]_{i}; followed by a sustained elevation of [Ca^{2+}]_{i}, was also observed when HEK293 cells expressing human MC1 (hMC1), hMC3, or hMC5 receptors were stimulated with the superpotent synthetic analog of α-MSH, NDP-α-MSH (17). In contrast to our data that compare activation of this pathway by two naturally occurring melanocortin peptides, no dose response curves or EC_{50} values were
A melanocortin receptor-mediated intracellular signaling pathway different from that presented in this report has previously been described. Konda et al. (18) demonstrated that MC3-R stably expressed in Hepa, L, or Chinese hamster ovary cells coupled to a signaling pathway in response to α-MSH, which resulted in a transient increase in inositol phosphates but no change in [Ca^{2+}]_i. Furthermore, when the PKA signaling pathway was inhibited in these cell types, activation of MC3-R by melanocortin peptides produced a sustained rise in [IP_3] at ≥10^{-11} M α-MSH and an increase in [Ca^{2+}]_i at ≥10^{-8} M α-MSH. This observation clearly differs from the response we have observed in HEK293 cells, where as little as 10^{-10} M α-MSH increases [Ca^{2+}]_i in the absence of an increase in inositol phosphates.

Mouse agouti protein is a competitive antagonist of mMC1-R coupling to a calcium signaling pathway in response to either α-MSH or desacetyl-α-MSH. Furthermore, agouti protein antagonizes both α-MSH and desacetyl-α-MSH coupling mMC1-R to the Ca^{2+} and PKA signaling pathways similarly (27). In contrast to this finding for the MC1-R, we have previously shown agouti antagonism of mMC4-R coupling to the PKA signaling pathway not to be competitive (maximum response is decreased) and also to be more potent in the presence of desacetyl-α-MSH than with α-MSH (27). It is of interest, therefore, that agouti protein also influences the mMC4-R coupling to the calcium pathway differently from its interaction with the MC1-R and also differently from the way it antagonizes MC4-R coupling to the PKA pathway. First, mouse agouti protein alters both α-MSH and desacetyl-α-MSH coupling mMC4-R to increases in [Ca^{2+}], but is more effective with α-MSH. Second, although mouse agouti protein decreases the sensitivity of melanocortin peptides coupling the MC4-R to the calcium signaling pathway, there is a dose-dependent trend for it to increase maximum responsiveness. It is unclear from our data, therefore, whether agouti protein is a competitive antagonist, or an enhancer, of the MC4-R coupling to the calcium signaling pathway.

The phenotypes of the e recessive (31) and MC4-R knockout (15) mice strongly support agouti protein antagonism of α-MSH coupling to the MC1-R inhibiting pigmentation, as well as agouti protein antagonism of desacetyl-α-MSH or α-MSH coupling to the MC4-R resulting in obesity and type 2 diabetes. An additional role for agouti protein driving the obesity, hyperinsulinemia phenotype in the yellow obese mouse has been proposed: one in which agouti protein stimulates calcium signaling independent of melanocortin receptor antagonism (17, 37, 38). This agouti protein stimulation of calcium signaling is clearly a mechanism different from the melanocortin peptide-stimulated rise in [Ca^{2+}], reported in this study.

The mechanism of melanocortin receptor antagonism responsible for the yellow obese mouse phenotype is believed to be agouti antagonism of MSH peptides coupling the MC1-R and MC4-R to increased cAMP with fivefold and three- to fivefold shifts, respectively, in their EC_{50} values (27). We show here that melanocortin peptides also couple the MC1-R and MC4-R to a second intracellular signaling pathway that results in elevations in [Ca^{2+}], the physiological consequences of agouti protein-induced six- and threefold shifts in EC_{50} values for α-MSH coupling MC1-R and MC4-R,
respectively, to increased [Ca\textsuperscript{2+}]; are currently unknown, but the magnitude of the shifts in EC\textsubscript{50} are similar to those for agouti antagonism of melanocortin peptides increasing cAMP. Therefore, the physiological relevance of the transduction of a calcium signal by melanocortin peptides and melanocortin receptors, as well as their interactions with agouti protein, requires further investigation.

We are grateful to Professor Michael Hanley, UC Davis, for helpful discussions, to Professor Garth Cooper, University of Auckland, for the use of the Hitachi F-4500 dual-wavelength spectrofluorometer, and to B. Stickney for technical support.

K. G. Mountjoy is supported by a Wellcome Trust Senior Research Fellowship of New Zealand, Health Research Council of New Zealand, National Child Health Foundation, and the New Zealand Lottery Board.

Present address of W. O. Wilkison: Zen-Bio, PO Box 12503, Research Triangle Park, NC 27709.

REFERENCES


