Gene expression profiling of gilthead sea bream during early development and detection of stress-related genes by the application of cDNA microarray technology

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Gene expression profiling of gilthead sea bream during early development and detection of stress-related genes by the application of cDNA microarray technology. Physiol Genomics 23: 182–191, 2005. First published July 26, 2005; doi:10.1152/physiolgenomics.00139.2005.—Large-scale gene expression studies were performed for one of the main European aquaculture species, the gilthead sea bream Sparus auratus. For this purpose, a cDNA microarray containing 10,176 clones from a cDNA library of mixed embryonic and larval stages was constructed. In addition to its importance for aquaculture, the taxonomic position and the relatively small genome size of sea bream makes it a prospective model for evolutionary biology and comparative genomics. However, so far, no large-scale analysis of gene expression exists for this species. In the present study, gene expression was analyzed in gilthead sea bream during early development, a significant period in the determination of quantitative traits and therefore of considerable interest for aquaculture. Synexpression groups expressed primarily early and late in development were determined and were composed of both known and novel genes. Furthermore, it was possible to identify stress response genes induced by cortisol injections using the cDNA microarray generated. The creation of gene expression profiles for sea bream by microarray hybridization will accelerate identification of candidate genes involved in multifactorial traits and certain regulatory pathways and will also contribute to a better understanding of the genetic basis of fish physiology, which may help to improve aquaculture practices.

Sparus auratus; stress response

IN RECENT YEARS, much effort has been invested in developing genome resources. Continuous improvements in technology have allowed rapid progress in the functional interpretation of sequencing data and the investigation of gene expression. The development of technology in model species has paved the way for high-throughput studies at a molecular level of non-model organisms of evolutionary, environmental, or economic interest.

The gilthead sea bream Sparus auratus is an organism of both economic and evolutionary interest, and it is one of the main European aquaculture species along with the Atlantic salmon (Salmo salar), rainbow trout (Oncorhynus mykiss), oyster (Crassostrea gigas), and sea bass (Dicentrax labrax). In addition to a basic understanding of the biology and reproductive physiology of this species, well-characterized culture methodologies for its reproduction and rearing also exist. From an evolutionary viewpoint, the gilthead sea bream is of interest as it belongs to the Perciformes, an order that underwent an explosive radiation 60 million years ago and contains a large number of other teleost taxa.

DNA microarray technology has become one of the significant tools in large-scale gene expression studies. For non-model fish species in which sequence data are not extensively available, only a few large-scale gene expression studies exist. For example, in the gobey fish Gillichthys mirabilis, the common carp Cyprinus carpio, and the Atlantic salmon S. salar, large-scale gene expression has been explored, respectively, in response to hypoxia, cold, and bacterial infection (19, 20, 49). Microarrays have also been used to study variation in gene expression within and among populations of the genus Fundulus (43). In contrast, extensive work setting up gene expression studies for functional analysis has been carried out in model organisms like Drosophila melanogaster (15, 41, 55), Saccharomyces cerevisiae (23), the mouse Mus musculus (54), the frog Xenopus laevis (1), Caenorhabditis elegans (25), and the zebrafish Danio rerio (36, 52).

Here, we report the construction and use of a cDNA microarray for the gilthead sea bream containing 10,176 cDNA clones [several thousand of which are estimated to be unique expressed sequence tags (ESTs)], which were obtained from a mixed embryonic/larval cDNA library (51). In addition, we provide insight into the developmental program of sea bream by characterizing gene expression profiles for five independent stages of sea bream development: neurula (stage 1), hatching (stage 2), pectoral budding/eye forming (stage 4), mouth opening (stage 6), and eye pigmentation (stage 7). Three main groups of genes were identified by K means clustering. More detailed analysis within each of the three groups led to the definition of expression models of up- or downregulated genes during early development as well as genes up- or downregulated before the period of mouth opening. The cDNA microarray chip developed was further validated and used to characterize the gene expression profile in the kidney of juvenile gilthead sea bream after treatment with cortisol. Identification in the present study of several genes previously reported to be up- or downregulated under stress conditions and during development in other teleost species as well as in gilthead sea bream corroborated the data set generated.

The results significantly scale up the exploration of the genome of a commercially important species for which ge-
nome information is scarce and provide first insights into the processes during growth and differentiation of the gilthead sea bream as well as during stress conditions.

MATERIALS AND METHODS

(Note that in microarray experiments, the labeled cDNA to be hybridized is called “target” and the DNA samples spotted on the microarray are called “probes.”)

Microarray Construction

Sample collection. Embryonic and larval stages of gilthead sea bream were collected from the breeding stock of the aquaculture facility at the Department of Aquaculture of the Hellenic Center of Marine Research (Crete, Greece), snap frozen in liquid nitrogen, and stored at -80°C. Kidneys were collected from untreated juvenile gilthead sea bream (n = 4) and from fish (n = 4) that received for 72 h a coconut oil implant containing 10 mg/kg (fish wet wt) cortisol. Experiments were carried out at the University of the Algarve (Faro, Portugal) in accordance with national legislation for the welfare of animals. Experiments were conducted in two 125-liter cylindrical tanks supplied with a continuous flow of oxygenated seawater at 20 ± 1°C using juvenile gilthead sea bream (25 ± 3 g) adapted for 1 wk to the experimental conditions. One tank contained eight untreated fish (control), and the other tank contained eight cortisol-treated fish. At the end of the experiment, fish were removed from tanks and decapitated, and the kidneys rapidly removed and placed in RNalater (Qiagen) at -20°C. No mortality occurred in the control tank, but two fish died in the cortisol-treated tank.

Bacterial cultures. For microarray production, a cDNA library of mixed embryos/larvae of gilthead sea bream (51) was used. Colonies were obtained by mass excision using the Lambda UNI-ZAP XR cloning Kit (Stratagene; La Jolla, USA) following the manufacturer’s instructions and were plated onto a 24 × 30-cm agar plate using 3-μm glass beads (Sigma-Aldrich; Munich, Germany). The colonies were picked into 384-square well plates containing 95 μl of 100 mM of each dATP, dCTP, dGTP, and dTTP; and 12.12 l of T7, 1.5 l of T3 primer (20 μM), 1 l of Hogness bromide (10 mg/ml), and the concentration was determined by 10 mM EDTA for 15 min at 65°C and neutralized by adding 10 μl of 1 M HCl. To remove unincorporated AA-dUTP and free amines, the reactions were purified with Qiagen columns using phosphate wash buffer, and coupling to the Cy dye ester was performed for 1 h at 20°C. No mortality occurred in the control tank, but two fish died in the cortisol-treated tank.

Culture PCR. The total volume of one PCR was 50 μl and contained 43.5 μl of reaction mix (2 μl of 10× PCR buffer; 0.04 μl of 100 mM of each dATP, dCTP, dGTP, and dTTP; and 12.12 μl distilled H2O), 1.5 μl of T7, 1.5 μl of T3 primer (20 μM), 1 μl of Taq polymerase (5 U/μl), and 2.5 μl of bacterial culture. The thermostability utilized was 94°C for 1 min 20 s, followed by 35 cycles of 94°C for 1 min, 50°C for 1 min, 72°C for 1 min 30 s, and a final step of 72°C for 5 min. PCRs were performed in duplicate to decrease the failure rate of reactions; duplicates were pooled, separated by agarose gel (2%) electrophoresis, and visualized by transillumination in the presence of ethidium bromide to check the insert size. Purification of PCR products was performed using the GFX-96 PCR Purification Kit (Amersham Pharmacia) according to the manufacturer’s instructions. In brief, 300 μl of binding buffer were added to the PCR products, mixed thoroughly, and transferred to the wells of the GFX-96 plate. The assembled plates were centrifuged at 1,800 g for 2 min, the liquid in the wash plate discarded, and the GFX-96 plate was reassembled on the top of the same wash plate. A second wash was conducted by adding 400 μl of wash buffer to each well of the GFX-96 plate and centrifuging for 5 min at 1,800 g; wash liquid was then discarded, and the GFX-96 plate was centrifuged for a further 30 s at 1,800 g to remove the remaining wash buffer. To recover the PCR products, the GFX-96 plate was assembled on the top of the sample collection plate, 50 μl of elution buffer were pipetted into the center of each well, and samples were incubated for 1 min and centrifuged at 1,800 g for 2 min. The purified products were dried under vacuum, reeuluted in 3× SSC and 1.5 M betaine to a volume of 10 μl, and transferred back to 384-well plates.

Spotting. Three rounds of spotting were performed: one round of 192 spots to test hybridization and spotting conditions, one round of 3,456 cDNA clones spotted in duplicate, and finally a third round of 10,176 cDNA clones again spotted in duplicate.

Test slides were spotted with the Affymetrix 417 Arrayer configured with 12 pins and rings. The final spotting was performed with the OmniGrid 100 robotic system (San Carlos, CA) configured with 48 pins (TeleChem SMP3; Sunnyvale, CA).

In total, 100 GAPII glass slides were spotted, with each glass slide contained 10,176 PCR-amplified inserts of the clones picked from the embryonic/larval cDNA library. The amplified cDNA products were spotted randomly and in duplicate in two separate subarrays, such that duplicate probe spots were not in the same quadrant but the next but one. As a control for hybridization and background, only buffer was spotted at the end of each spotting group. The spotted glass slides were baked at 80°C for 2 h to cross-link the DNA onto the glass slide. The slides were then incubated in blocking buffer at 50°C for 30 min, washed twice with Milli-Q water at room temperature for 2 min, and dried by spinning in an Eppendorf 5890R centrifuge for 7 min at 800 rpm.

Data Acquisition

RNA extraction. Total RNA was extracted from five larval stages (neurula, newly hatched, pectoral budding, open mouth, and eye pigmentation stages) and also from cortisol-treated and untreated juvenile gilthead sea bream kidneys using the mini-RNAeasy extraction kit (Qiagen). The quality of the total RNA obtained was checked by gel electrophoresis on a 1% agarose gel containing 0.05% ethidium bromide (10 mg/ml), and the concentration was determined by spectrophotometry.

Labeling. Ten micrograms of intact total RNA were directly labeled following the TIGR protocol (http://www.tigr.org/tdb/microarray/protocols/TIGR.html) with the red fluorescent dye ester cyanine 5 (Cy5) and green fluorescent dye ester cyanine 3 (Cy3) (Amersham Pharmacia). In brief, cDNA was synthesized by incorporating aminooallyl dUTP (AA-dUTP). Synthesis took place at 42°C for 3 h. RNA was hydrolyzed with 10 μl of 1 M NaOH and 10 μl of 0.5 M EDTA for 15 min at 65°C and neutralized by adding 10 μl of 1 M HCl. To remove unincorporated AA-dUTP and free amines, the reactions were purified with Qiagen columns using phosphate wash buffer and phosphate elution buffer. The purified sample was dried in a Speed Vac. AA-cDNA was resuspended in 0.1 M sodium carbonate buffer, and coupling to the Cy dye ester was performed for 1 h in the dark at room temperature. The removal of uncoupled dye was performed using the Qiagen PCR Purification Kit. Finally, samples were tested for successful dye ester incorporation by spectrophotometry. The labeling was considered successful when total dye incorporation was over 200 pmol and the ratio of nucleotides to dye molecules was <50.

Hybridization. The two targets (Cy3 and Cy5 labeled) were mixed, and the volume was increased to 500 μl with 1× Tris-EDTA buffer (TE). The mixture was applied to a microcon 50 spin filter column and centrifuged for 13 min at 8,000 rpm to reduce the target volume to 15 μl. To the target DNA, 2 μl of 10 mg/ml herring sperm DNA were added, and the volume was increased to 15 μl with 1× TE. The target was denatured for 2 min at 95°C, incubated on ice for 1 min, and finally left at room temperature for 5 min. An equal volume (15 μl) of hybridization buffer was added to the target; the solution was vortexed, spun down, and carefully pipetted onto a coverslip; and the microarray was inverted and placed in a hybridization chamber containing a prewetted Whatman paper strip. Hybridization took place overnight in a water bath at 42°C.
Washing. After hybridization, the chip was carefully taken out of the chamber, and the coverslip was removed by immersion in 4× SSC. The chip was then washed for 5 min in wash buffer 1 (2× SSC, 0.1% SDS, and Milli-Q water), 5 min in wash buffer 2 (0.1× SSC, 0.1% SDS, and Milli-Q water), and twice for 2.5 min in wash buffer 3 (0.1× SSC and Milli-Q water). All washing steps were carried out at room temperature with vigorous shaking. Finally, the slides were quickly dipped in 0.05× SSC and in Milli-Q water. Drying of the slides was performed by centrifugation at 1,000 rpm for 12 min in a 50-ml Falcon tube at room temperature. All the above steps were performed in the dark.

Data processing and analysis. Microarray gene expression data for embryonic and larval stages were obtained by comparing the results of each stage with each other using a loop design (Fig. 1). All experiments were performed by dye swap hybridizations of the sea bream cDNA microarray chip. Taking into account all possible combinations, 20 hybridizations were performed in total within this experimental setup.

For the study of treated versus untreated kidneys, a total of four hybridizations was performed. The labeling of total RNA from the kidney was performed using a sample from one individual. For each cDNA microarray chip, two series of digital images were produced by dual laser scanning with a GMS 418 Array Scanner (MWG). The first series was obtained by illuminating the array at 635 nm (excitation of Cy5), and the second series was obtained by illuminating the array at 532 nm (Cy3). For both illuminations, three images were obtained from each slide with two different signal settings of the photomultiplier tube (gain and light amplification) and two settings (90% and 100%) of the laser power were used. All images were captured in TIFF format.

Spot finding, image segmentation, and quantification was carried out with ImaGene 5.0 microarray image analysis software (BioDiscovery; Los Angeles, CA). Briefly, a grid was created taking into account the number of spots, the print pattern, and the size of the spots. Spot finding was performed using a semiautomatic method in which each spot is checked for proper alignment within the grid manually, as this gives more accurate data collection than fully automatic methods. Finally, spot quantification was performed to obtain the expression level of each gene deposited on the array. For downstream analysis, Cy3 and Cy5 scans were chosen according to the ratio of the Cy3-to-Cy5 ratio after spot quantification (ImaGene 5.0 software package). A ratio of around 1 is expected because the majority of genes do not change expression.

Data set preparation and quality control. The quantified data were analyzed using the software program GeneSight lite included within the ImaGene 5.0 software package. For each data set, the background was subtracted, and poor, empty, or negative spots (see the ImaGene 5.0 manual) were flagged as unreliable and were excluded from downstream analysis. For each data set, the ratio was calculated and normalization was performed using the z transform logarithm of GeneSight lite. The normalized data were stored in a relational database using FileMaker Pro5 software (FileMaker; Santa Clara, CA). Genes selected as being differentially regulated were those in which the ratio of experiment to control was at least 2σ away from the mean value obtained.

The correlation of measurements was determined using the results for duplicate spots on the array.

Raw and normalized data were submitted to ArrayExpress, a public repository for microarray gene expression data at the European Bioinformatics Institute (experiment: E-MEXP-181, array: A-MEXP-110), as well as to the National Center for Biotechnology Information (NCBI) Omnibus under Accession Nos. GSE 2064 and GSE 1887; all clones are available on request. All sequence data have been submitted to the NCBI EST database (Accession Nos. CB184056–CB184594 and CV133223–CV133736).

Analysis. Normalized data were further analyzed using K means clustering and univariate analysis with the statistical software package RSPS 12.0 (SPSS; Chicago, IL).

The K means clustering method is the most appropriate clustering method when analyzing populations of more than 200 data points and was used to group all data points obtained setting cluster numbers from 2 to 5. Distances were computed using simple Euclidean distance. The cluster-cluster distance was calculated by determining the distance between centroids. Expression patterns of genes over the course of the developmental stages were obtained by univariate analysis using the statistical package RSPS.

RESULTS

Microarray Construction

As a first approach toward gene expression studies in sea bream, a cDNA microarray of an embryonic/larval cDNA library was constructed. First, preliminary cDNA microarrays were constructed with 192 cDNA clones spotted 5 times each of genomic sea bream DNA were included as controls. Hybridizations were carried out with the following targets: 1) labeled 50-mer oligonucleotides designed from previously sequenced gilthead sea bream clones, 2) labeled PCR products of gilthead sea bream dlx3 and dlx6, 3) gilthead sea bream ribosomal protein L13a, 4) zebrafish nacre, 5) zebrafish dlx3, and 6) labeled total RNA extracted from two different larval gilthead sea bream stages. The zebrafish targets hybridized only with the spotted probes from zebrafish, indicating the stringency of the hybridization conditions utilized. Cross-hybridization between gilthead sea bream cDNA probes and the zebrafish target was not observed. On the basis of these preliminary tests, a final cDNA microarray containing a total of 10,176 cDNAs was printed in duplicate onto aminosilane-coated slides from Corning. Of the 10,176 spotted cDNA clones, 768 had previously been sequenced (51). It is expected that several thousand different cDNA clones are represented on the chip, as around 75% of the previously sequenced clones were unique. The correlation, determined before normalization, between duplicate spots used for downstream analysis in each hybridization subsequently was found on average to be 85%. The quality of the array was indicated by the high $R^2$ values obtained after univariate
analysis of genes represented more than once in the array (e.g., Fig. 6B) and by the comparable expression results of known genes in the present study and previously published studies.

Isolation of Candidate Genes Involved in Development

To obtain an overview of genes involved in sea bream development, targets were chosen that represented five time points covering different developmental events from neurula through to pigmentation of the eye (Fig. 2). A group of cDNA clones out of each hybridization, for which z scores of $x^2/H_11349$ or $x^2/H_11350$ were detected, were picked, sequenced, and analyzed using BLASTN and BLASTX searches against the NCBI nr database. Sequence alignments resulting from the BLASTN search against the nr database were examined, and a sequence identity of $65\%$ over a minimum range of 80 bp and an E value of $10^{-5}$ were chosen as the criteria for designation of putative gene identity (cf. Supplement A; available at the Physiological Genomics web site). When the minimum alignment was <80 bp, BLASTX analysis was also performed to confirm the BLASTN result. Some cDNA clones with a low BLASTN e value and no hit in BLASTX were also selected as they shared a similar expression pattern, also showing the quality of the hybridizations [e.g., SA-E33D_A08, SA-E42D_C10, SA-E60B_A11, and SA-E60C_B11 (all clones assigned to $b$) AU301079.1 C. carpio cDNA clone: 3-033, expressed in barbel]. Subsequent sequence analysis revealed that the four cDNA clones encoded the same gene, and, in BLASTX, the assembled sequence shared sequence similarity with lipocalin-type prostaglandin D synthase-like protein ($D. rerio$). The above procedure was performed for all clones with a low e value and a similar expression pattern which by BLASTN were assigned to a specific clone.

Frequently used methods for grouping genes by their expression patterns over the experimental variables are hierarchical clustering and K means clustering. In this study, K means clustering was performed on the data points obtained after z score normalization, as this is more suitable for analyzing populations of more than 200 genes (21). Clustering of all sequenced clones on the chip [clones already known from Sarropoulou et al. (51) and clones selected by z score values] before the selection step by microarray hybridization revealed the largest F values throughout the 10 experiments when the cluster number was set to 3, indicating the greatest separation between clusters (Fig. 3) (SPSS). When only genes showing significant differential expression using the z transform method in at least 1 of the 10 experiments were taken into consideration, the clusters were more clearly defined.

Fig. 2. Pictures of gilthead sea bream larvae at the significant time points during development that were chosen for transcription profiling.

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1 The Supplemental Material for this article (Supplements A and B) is available online at [http://physiolgenomics.physiology.org/cgi/content/full/00139.2005/DC1](http://physiolgenomics.physiology.org/cgi/content/full/00139.2005/DC1).
account, K means cluster analysis again revealed the largest F values when the cluster number was set to 3 (Table 1 and Fig. 3).

The three clusters represent, respectively, genes that are mainly expressed early in development, genes mainly expressed in later developmental stages, and genes showing differential regulation in the middle of the stages selected for hybridization. Expression profiles were visualized using the software TreeView (12), which plots the experiments on the y-axis and the differentially expressed genes on the x-axis. The colors displayed indicate whether the gene is up- or downregulated. In this study, as shown in Fig. 4, the green color indicates that genes are downregulated in the later developmental stages considered and upregulated in the earlier developmental stages.

Isolation of Candidate Genes Involved in Stress Responses

Microarray experiments were performed with kidneys of cortisol-injected fish versus control fish to identify known and novel genes potentially involved in the stress response induced by increased circulating cortisol levels. These experiments revealed 257 significantly regulated genes, including several genes related to carbohydrate metabolism as well as genes involved in Na+/K+ transport, polyamine biosynthesis, and iron homeostasis. In total, 206 clones were sequenced, and 35% did not show a significant BLAST hit (Supplement B).

**DISCUSSION**

In the present study, insight into the developmental program of the gilthead sea bream is established by the construction and application of a cDNA microarray from embryos and larvae of the gilthead sea bream. Furthermore, this platform was also validated for use in studies of the stress response in gilthead sea bream.

To identify genes involved in the development of sea bream, five different developmental stages (neurula, hatching, pectoral budding/eye development, mouth opening, and eye pigmentation) were chosen for the first round of experiments. Microarray studies providing specific gene expression patterns have been widely used to isolate genes and as a sensitive indicator of gene function (for a review, see Ref. 57). Most of the microarray experiments use an indirect design (also called reference design) as it has the advantage of allowing easy analysis and interpretation without the need for extensive statistical tools. However, if subtle variations between the experiments are of interest, then a design with direct comparison (also called a balanced or loop design) is preferable (56) as it allows comparison of experiments with much greater precision (26, 27). In the present study, a direct experimental design (Fig. 1) for the five different developmental stages of the gilthead sea bream (Fig. 2) were used. To corroborate microarray data, it is desirable to confirm the specificity of a microarray chip as well as the hybridization and analysis method used by analyzing the expression profiles of known genes (50). In this context, several approaches were taken, including comparison of expression profiles with previous studies in other fish species and comparison from a functional genomics viewpoint of the results for developmental stages with those of cortisol-injected fish as well as specific functional studies of genes found in the literature. As expected, genes encoding the translation machinery (according to the gene ontology classification of cellular components) were only scantily represented among the significantly regulated genes in the experiment, which is in direct contrast to results previously obtained with an EST-based study of the same library (51). Similar microarray experiments performed for developmental stages of zebrafish (36, 52, 53) indicated the increased expression of genes involved in myogenesis at the 24-h stage, corresponding to the period of pharyngula, and the 72-h stage, corresponding to the hatching period. These results are congruent with the present results in which there is an upregulation of genes encoding myogenic proteins at stage 2, when hatching occurs (Fig. 5). The results obtained from our stress response

![Fig. 3. Canonical discrimination functions. A and B: clustering of all sequenced clones (A) and clustering of clones with differential expression identified via a z score of z < −2 or z > 2 by the K means method (B). The data points given (cases 1–3) were grouped in three main clusters (colored blue, green, and gray, respectively). The group centroid represents the average value of the cases contained in each cluster.](http://physiolgenomics.physiology.org/)

<table>
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Largest F values (>100) are indicated in bold.
hybridizations (discussed further below) also corroborate the microarray data for developmental stages. In addition, bibliographic corroboration of the data obtained in this study was also found, e.g., a gene belonging to the enolase superfamily was upregulated in the stress microarray study, as reported in a previous study (48) of \( \alpha \)-enolase expression under stress conditions. Furthermore, as a nonnormalized cDNA library was spotted, several genes were represented as multiple duplicates on the same chip, and the results showed good reproducibility of the up- or downregulation of specific genes after clustering and applying statistical analysis by univariate analysis of variance (21).

Cluster Analysis

Cluster analysis revealed three main groups of genes with stage-specific expression. \( \text{Cluster 1} \) contains genes showing upregulation in the last two stages as well as genes showing upregulation in stage 4, with a sustained high level of expression in the later stages. Among the genes showing upregulation in the last two stages, stage 6 (mouth opening) and stage 7 (eye pigmentation), many candidate genes for visual pigmentation were found, which is in line with phenotypic observations of eye pigmentation [168 hours postfertilization (hpf)] (Fig. 6). In contrast, many of the genes that are found to be upregulated in

Fig. 4. Heat map images showing the genes grouped by the \( K \) means clustering method in \( \text{cluster 1} \) (genes expressed mainly in the middle and later ones of the selected developmental stages), \( \text{cluster 2} \) (genes expressed mainly in the early ones of the selected developmental stages), and \( \text{cluster 3} \) (genes up- or downregulated transitorily in the middle of the selected developmental stages). Stages are indicated above each column. Gene expression is shown in rows. The quantitative changes in gene expression are represented in color: red indicates upregulation in the later stage, whereas green indicates upregulation in the earlier stage of the hybridization experiment.

Fig. 5. Genes involved in myogenesis, showing upregulation at stage 2, when the larva has just hatched. On the x-axis, the stages are plotted, and, on the y-axis, the estimated marginal means are plotted. The univariate analysis of variance for randomized design uses the criterion of the multiple \( R^2 \) statistics measure. If all the independent variables fit on the regression line, then \( R^2 \) is >0.5.
stage 4 encode metabolic enzymes, studies of which are of special interest for fish physiology and aquaculture.

In fish larvae, nutrients are initially supplied only by the yolk reserve, although at mouth opening and the onset of exogenous feeding nutrients are absorbed across the rudimentary digestive tract. A major problem for aquaculture is the low acceptance of artificial diets by larval fish. Large variations in ingestion rates are observed in feeding experiments, and, in some cases, first-feeding marine fish larvae ingest artificial feed but fail to grow. Advances in this area have produced better results (9, 13, 29, 31, 34), but, in most species, problems arising from the use of formulated feeds at the onset of exogenous feeding still persist. The larvae of gilthead sea bream hatch from pelagic eggs, and full development of the digestive system only occurs at metamorphosis (40). Determination of the onset of enzyme production is therefore essential for choosing appropriate larval diets and establishing the earliest possible time for weaning.

In the present study, many genes encoding metabolic enzymes including amylase are upregulated before mouth opening, namely, in stage 4 (Fig. 7), which corresponds to 4 days posthatched. At this time, the pectoral fin is budding and the eye is beginning to form. Studies of the amylase gene in the winter flounder Pleuronectes americanus show that the onset of amylase expression is at a similar stage, 5 days posthatched (10). Interestingly, in the present study, both fructose-1,6-bisphosphatase and fructose-1,6-bisphosphate aldolase B, enzymes involved in gluconeogenesis, are downregulated at stage 4, just when amylase is upregulated (Fig. 7). This may indicate a concerted switch from gluconeogenesis, when nutrients come from yolk lipids, to the glycolytic pathway as soon as exogenous carbohydrates become available. Although amylase activity in D.
monitor the acceptance of artificial food to larval fish.

during the development of the digestive system and help to

Furthermore, microarray analysis of feeding experiments

knowledge about the metabolic capabilities of different larval
development, and, in particular, peptidases, will generate

ing of further genes encoding digestive enzymes throughout
development (18, 48). Genes encoding proteins involved in Na+/K+
transport were also found to be upregulated, which is in

concordance with studies showing that cortisol stimulates

branchial Na+/K+-ATPase activity, an enzyme crucial for

osmoregulation (37).

A number of genes encoding proteins of interest were found
to be up- or downregulated under stress conditions in the

present study. One such transcript encodes ornithine decarbox-
ylase (ODC), a key enzyme of polyamine biosynthesis that is
elevated in rapidly proliferating cells. Overexpression of ODC
has been associated with carcinogenesis in mammals (4, 39).
ODC is also a marker of liver neoplasia in winter flounder (P.
americus) (32), and its upregulation in stressed gilthead sea
bream may cause a predisposition to tumors, an issue of some
importance for aquaculture. Interestingly, two of the main
proteins in iron metabolism, ferritin heavy and light subunit
and the transferrin receptor, were downregulated by cortisol.
Oxidative stress and iron metabolism are interrelated in mul-
tiple ways. Ferritin is an intracellular molecule that stores iron
in a soluble nontoxic form and was found to be induced in
response to hypoxia in a gene expression profiling study in G.
mirabilis (18) and repressed in response to quinone-induced
oxidative stress in a study of murine B6 fibroblasts (17).
The transferrin receptor is an iron transport protein that was like-
wise found to be repressed in murine fibroblasts in response to
oxidative stress (17). Because it is known that intracellular iron
can be a rate-limiting factor for cell growth and proliferation
(35), a downregulation of intracellular iron levels may be
associated with the suppression of growth observed under
stress conditions. Furthermore, elevated levels of transferrin
receptor were found in Atlantic salmon macrophages activated
by infection with Piscirickettsia salmonis (49), suggesting that
its downregulation could contribute to immunosuppression.
In humans, transferrin is used as a clinical marker as its concen-
tration level correlates with the total mass of immature ery-
throid cells (47). Potentially, it could also be employed as a
diagnostic marker in aquaculture. For this purpose, it will be
important to determine whether the decrease of the transferrin
receptor seen during stress primarily reflects a decrease in the
number of hematopoietic cells or a lowering of intracellular
iron levels.

Another area to be explored in future work is the use of
whole fry as an effective sample of stress response as well as
the effect of stress on specific developmental stages of sea
bream as several stress-responding genes isolated in this study
were also found to be regulated during development.

In conclusion, the present study provides a first glimpse of
how the global analysis of gene expression, using material
from appropriately chosen conditions or tissues as a target, can
yield insight into the molecular basis of development and stress
in aquaculture species. The data set presented here will be a
basis for future expression profiling work in this area. It has
been proposed that genes with similar biological function
constitute synexpression groups with coordinated expression
throughout development (16, 41), an expectation that is borne
out by our clustering studies, e.g., of proteins encoding for
epithelial tissue. This opens up the possibility of focusing
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Lazo JP, Dinis MT, Holt JG, Faulk C, and Arnold CR. Co-feeding microparticulate diets with algae: towards eliminating the need of zoo-

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